



## **Occurrences and frequency of fungi isolated from fast foods and spices**

**Hager A. Bashir, Amira E. Sehim, Rasha Y. Abdelghaffar, Mahmoud M. Amer and Amany M. Emam**

Botany and Microbiology Department, Faculty of Science, Benha University, Benha, Egypt

### **Abstract**

Meat products especially beef, luncheon and burgers are one of the most popular meals in many countries in the world including Egypt, which were found to be highly contaminated with fungi especially *Aspergillus* and *Penicillium* spp during the manufacturing process leading to high economic losses. Also, food spoiling has a public health hazard due to the production of their mycotoxins. Therefore, the current study concentrated on the isolation and identification of mycotoxigenic fungi that were connected to the samples of processed meat. So, a total of 54 fast food samples, including corn flex, meat spices, luncheon, burgers, indomie, meat, sausage, crisps, karate (snacks), biscuits, maize and soebean, were gathered from various locations in the Qalubia governorate in Egypt. The collected samples were examined mycologically. According to the findings of this study, the luncheon samples under examination had the highest total fungus count with 313 fungal colonies/10g of the samples, followed by, meat spices (153) and crisps (152) fungal colonies/10g. While indomie, sausage and soebean showed the lowest mean fungus count with (14, 25 and 25, respectively) and the *Aspergillus niger* was the most frequently and counted 248 in luncheon, 98 in basterma and 86 in burger followed by *A. flavus* 85 in Crisps, 70 in corn and 66 in meat spices. Because of this, a lot of meat products and spices have severe fungal contamination. These findings indicate the risk of fungal contamination exposure to consumers due to the high consumption of fast foods and spices which may be susceptible to fungal infection, leading to mycotoxin contamination if the storage conditions are favorable for fungal growth.

**Keywords :** meat products, spices, *Aspergillus niger*, *Aspergillus flavus*

### **1- Introduction**

Ready-to-eat (RTE) products, such as luncheon, basterma and hawawshi are prepared to be eaten

without needing cooking and therefore often consumed without additional cooking steps. Post-process handling is a cause of recontamination of RTE meat products especially with food pathogen.

Consumers may choose to cook them for a better taste or appearance[1]. Some ready-to-eat foods were considered as potentially hazardous, because such foods can support the growth of microbes. Such food must be kept at certain temperatures and conditions to decrease the growth of any microorganisms that may be found in the food or to prevent toxins formation in the food. Due to the nature of these foods and their methods for preparation involving extensive handling, they were contaminated during storage, distribution facilities, soil, water, air environment and human activities including the food handlers and vendors[2]. Meat products provides an excellent growth media for a variety of microorganisms[3].

It is believed that there is a big problem with meat products that contaminated with several types of fungi because it makes decay and disintegration more likely, it has an effect on the meat quality products. The most important side about the fungal spoilage of food is, the formation of mycotoxins especially aflatoxins, which considered the main toxic secondary metabolites of *Aspergillus* spp such as *A. parasiticus*, *A. flavus* and *A. nomius*[4,5].

Some fungi especially *Aspergillus* spp. had a bad effect on the human health; it not only causes diseases, but also contaminated the human diets. Some mold species such as *A. flavus* and *A. parasiticus* are toxigenic and produce aflatoxin in foods[6,7]. Since meat products provide a significant amount of (proteins, essential amino acids, fats, minerals, vitamins, and other nutrients) they are more appealing of highly nutritious diets for human consumptions. They are thought to be the best culture media for the development of many organisms. However, due to their high levels of

moisture, large amounts of nitrogenous substances, abundant mineral supply, presence of some fermentable carbohydrate (glycogen) they were an ideal media for the majority of microbes[8].

Therefore, the growth of some fungal species is dangerous, because they can produce several mycotoxins[9].

Mycotoxin is a problem in the food industry that has an effect on human and animal's health. In storage condition, fungal bio-deterioration of stored food is a chronic problem especially in tropical hot and humid climates due to excretion of mycotoxins [10,11], that can be produced by different fungi such as *Aspergillus*, *Penicillium*, *Alternaria*, *Fusarium*, *Cladosporium*, *Mucor*, and *Rhizopus*[12]. Some fungi, including *Aspergillus*, *Fusarium*, *Penicillium*, and *Alternaria* have the capacity to create mycotoxins, which harmful byproducts, that can contaminate food under specific conditions[13,14].

According to estimates, more than five billion individuals contaminated food every day and are exposed to mycotoxins through unidentified pathways every day[13]. The disease caused by ingestion of Mycotoxin called Mycotoxicosis [15].

The aim of this research is to evaluate certain (ready-to-eat) food items available in Qalubya shops in terms of their mycological quality. This mycological study investigates the total fungal counts; isolation and identification of recovered fungal species.

## 2. Material and Methods

### 2.1. Samples collection

About (54) samples of different meat products and most consumed corn products also different spices

were collected from several markets in Qalubya Governorate. The food samples from supermarkets were taken to microbiology lab in faculty of science, Benha University in sterile plastic bags in Ice-Box, according to Chessbrough [16].

Using established techniques described in ICMSF [17], the samples were examined for bacteria and fungi related to human health.

## 2.2. Sampling preparation

90 ml of 1% peptone water was added to an aseptically prepared blender jar containing 10 grams of each sample. The mixture was then homogenized for two minutes in the sterile warring blender before being diluted ten times in serial fashion. The previously made serial dilutions were inoculated separately in one millimeter portions into petri dish plates and combined with potato dextrose agar medium.

The plates were combined, given time to firm up, and then incubated for 5-7 days at 25°C.

## 2.3. Isolation and purification of fungi

Transferring a single fungal colony to fresh medium plates on potato dextrose agar resulted in pure cultures of colonies [18].

On PDA agar plates, fungal colonies were also cultivated for roughly 7 days at (25°C).

## 2.4. Cultural and morphological

Based on cultural and morphological characteristics on particular media and available literature; slides were continuously examined under multiple powers of microscope, namely 10 and 40X. These

characteristics were then compared with the description provided by another study [19] for the type *Aspergillus*, the isolates of *penicillium* spp. were detected according to Ramirez [20] and Pitt [21]. All developing fungus were cultivated on PDA slants and kept in refrigerator for the genera of imperfect fungi. The formula was used to determine the frequency of fungi and the proportional percentage of each species within a genus of fungi [22].

$$\text{Frequency\%} = \frac{\text{Number of samples infected with fungi} \times 100}{\text{Total number of sample analysis}}$$

$$\text{Relative Percentage \%} = \frac{\text{Number of fungal species isolated} \times 100}{\text{Total number of fungi}}$$

In Petri dishes with Czapek's yeast extract medium (CYA), fungus isolates were cultivated [23]. The media was consisting of (g/L): Sucrose, 30; Na<sub>2</sub>NO<sub>3</sub>, 2; K<sub>2</sub>HPO<sub>4</sub>, 1; KCl, 0.5; MgSO<sub>4</sub>.7H<sub>2</sub>O, 0.5; FeSO<sub>4</sub>, 0.01; ZnSO<sub>4</sub>, 0.01; CuSO<sub>4</sub>, 0.005; yeast extract; chloramphenicol, 0.25 and agar. Cultures were incubated at 28° C for 7 days. The growing fungi were identified using the colony's features (growth rate, color, texture, and reverse pigmentation) and microscopic information (shape of conidiophores, conidiogenous cells, and conidial dimensions). For easier visualization, lactophenol cotton blue was used to dye fungus hyphae and conidia.

The stain contained 20 g; of phenol crystals (C<sub>6</sub>H<sub>5</sub>O<sub>4</sub>), 0.05g of cotton blue (or methylene blue), 40 ml of glycerol, 20 ml of lactic acid (CH<sub>3</sub>CHOH COOH), and 20 ml of distilled water.

Axiostar trinocular microscope from Zeiss, Germany, was used for microscopic investigation and imaging. A digital camera made in Japan with 7.1

megapixels, the Canon G6, is included with the microscope [24].

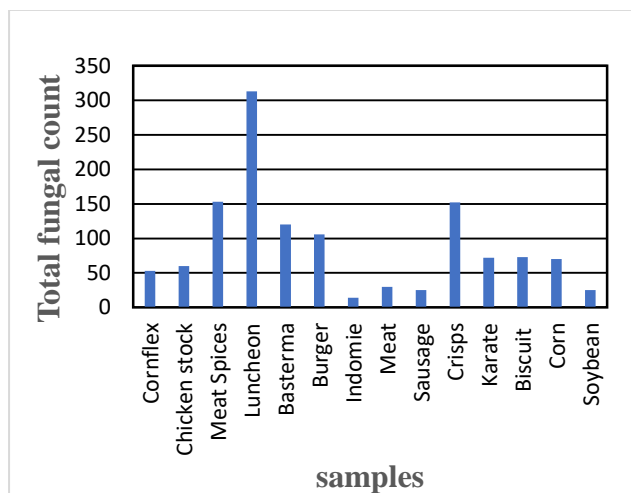
### 3- Results and Discussion

#### 3.1. The total fungal count of the analyzed processed samples

Data presented in Fig. (1) showed that mycological examination of fourteen processed samples, i.e. cornflex, chicken stock, meat spices, luncheon, basterma, burger, indomie, meat, sausage, crisps, karate (snacks), biscuit, corn and soebean. The greatest total fungal count was found in the luncheon samples, which noted 313 fungal colonies/10g: of the samples under examination, followed by, meat spices (153) and crisps (152) fungal colonies/10g while the least mean fungus count was found in samples of indomie, sausage and soebean, which recorded (14, 25 and 25, respectively) fungal colonies/10g. Similar results were found by many researchers [25].

According to their examination of 20 beef burger samples collected from the Assiut, Giza, and Cairo Governorates, the fungus population ranged from 164 to 528 colonies/g. The overall fungal count of the beef samples, according to Omorodion & Odu [26], ranged from  $(6.0 \times 10^4)$  CFU/g to  $(4.4 \times 10^5)$  CFU/g. The differences in fungal counts may be explained by the geographical location of producing companies, the length of time, these items were held, and the cleanliness of the personnel handling them. These findings support with [27], who found that the incubation temperature, types of media, and techniques of food analysis are all factors that affect the fungal counts. However, 92% of the samples of beef luncheon that identified as being contaminated with molds [28]. Besides [29] who found that in 70% of

the samples of hot dogs, mold was discovered. Different conditions for the relevant meat products (raw or cooked), the quality of the meat and its additives, sanitation during processing and packaging, maintenance of adequate refrigeration from the processor to the retail level and to the consumers, sanitation during handling at the retail stores, and lastly the laboratory technique for detecting fungi contamination tested can all be used to explain the varying percentages of fungi-contaminated samples [30]. By causing the breakdown of the meat's components and the release of various acids and gases, which changes the meat's odor and flavor, the presence of molds can lead to the spoiling of meat products. Some molds are capable of producing mycotoxins, which are hazardous by-products that can cause cancer [31].



**Fig.(1): Total fungal count of the examined processed samples.**

#### 3.2. Fungal frequency associated with tested samples:

A complete list of all mycoflora recovered from examined processed samples was provided in Table 2. This table's findings showed that seven fungal genera

were recognized and reported. These are *Aspergillus, Penicillium, Mucor, Rhizopus, trichoderma, Fusarium* and *Alternaria*. The *Aspergillus niger* and *A. flavus* were the genus most detected at high frequency in all samples. The most common species were *A. niger* 87%, *A. flavus* 59.3% followed by *Penicillium* spp. 25.9 %. Also, The highest relative density percentage was shown by *Aspergillus* species especially, *A. niger* were the most common and isolated with higher density (62.9%) followed by *A. flavus* (31.2%) In this study the fungal genera were recovered, known to be mycotoxigenic according to [32]. The status of meat products is affected by fungi that enter through meat spices, and other components as well as through the processing environment, equipment, and handlers[5].

The same results have been achieved by [33] which found the presence of *Aspergillus* spp. i.e. *A. niger* (40%) followed by *A. flavus* (27%) *A. ochraceus* (15%), *A. fumigatus* (10%), *A. japonicus* (5%) and *A. sclerotiorum* (3%) in corn.

*A. flavus* and *A. parasiticus* can contaminate corn while being transported and stored. Due to the fact that both species' spores have a lengthy air survival lifespan and the ability, to spread widely from one place to another [34,35].

On the other hand, the environmental conditions in the factories, warehouses, freezers, and stores are favorable for the molds. Although molds can develop inside the products, they do so more frequently on

the outside of various kinds of meat and meat products. Significantly to food spoiling, some molds can even produce mycotoxins that can be dangerous to people [36]. As well as spices have been used in many industries. They frequently have high levels of mold contamination. The most prevalent fungal species that contaminate spices are *Aspergillus* and *Penicillium* spp [37]. Additionally, [38] assessed the microbiological quality of meat products sold in a number of supermarkets and stores in the Gharbia Governorate, including luncheon, basterma, kofta, and burgers, as well as spices (used in meat processing). In 120 analyzed meat products and 33 samples of spices, they found 9 different mold genera, including *Aspergillus, Penicillium, Acremonium, Cladosporium, Geotrichum, Mucor, Claveolaria*, the main *Penicillium* species were *P. citrinum*, and *P. aurantigreu*. *P. chrysogenum* was the most frequently isolated species from burger samples, while *P. paxilli* and *P. restrictum* were the most frequently recovered species from kofta samples. Only samples from luncheon and basterma were found to have *p. citreognigrum* and *P. carneum*. The most fungi were found in the luncheon samples, then in the basterma samples, while hawawshi samples had lower mold counts. The fungus genera that were found belonged to the genus *Aspergillus, Penicillium, Cladosporium, Mucor, Eupenicillium, and talaromyces*. *A. niger*, followed by *A. flavus* and *A. parasiticus*, had the highest incidence of isolates among *Aspergillus* species., according to [39].

**Table (1) :Fungal frequency associated with examined samples.**

Total fungal species	Total No.of isolates	No.of positive samples	Frequency (%)	Relative density (%)
<i>A.niger</i>	790	47	87.04	62.99

<i>A.flavus</i>	395	32	59.26	31.20
<i>Aspergillusoryzae</i>	19	5	9.26	1.52
<i>A.ochraceous</i>	1	1	1.85	0.08
<i>Pencillium sp.</i>	33	14	25.93	2.63
<i>Mucar</i>	2	2	3.70	0.16
<i>Rhizopus sp.</i>	10	6	11.11	0.79
<i>Trichoderma sp.</i>	4	1	1.85	0.31
<i>Fusarium sp.</i>	5	2	3.70	0.39
<i>A.paraciticus</i>	2	1	1.85	0.15
<i>A.terrus</i>	3	2	3.70	0.23
<i>Alternaria</i>	2	1	1.85	0.15

### 3.4 .Occurrence of fungi isolated from the analyzed samples

The results in Table 2 showed that the samples that were infected with mycoflora were grown on PDA media. The total fungal count (TFC) was 1266 isolates. Data also revealed that *Trichoderma*, *Aspergillus*, *Pencillium*, *Fusarium*, *Rhizopus*, and *Alternaria* were the most commonly isolated fungus species. It was noticed that *Aspergillus* spp. (*A.flavus* and *A. niger*) were the most frequent in all samples. We noticed that *A. niger* was the most frequent and counted 248 in luncheon, 98 in basterma and 86 in burger followed by *A.flavus*. 85 in Crisps, 70 in corn and 66 in meatspices Also, *A.oryzae* isolates was examined in luncheon that recorded 12 isolates , the predominant *Penicillium* species isolated from cornflex, chicken stock, meat spices, luncheon , basterma, Burger. These results of fungal incidence accept with the results provided by [5,23,33,34]. They discovered that *Aspergillus* spp. and *Penicillium* spp. were the most often isolated fungus from meat products. As well as production areas, shops, and refrigerators all ideal environments for the growth of fungus on meat and meat products. Which they

were highly contaminated with mold especially *Aspergillus* spp and *Penicillium* spp which may gain access during the manufacturing process leading to high economic losses and have a public health hazard due to the production of mycotoxins that create a major economic losses and public health problems. These conclusions resemble those made by [5] who examined beef burgers, sandwiches kofta, oriental sausage, and basterma and discovered nine fungal genera. The genus *Aspergillus* had the highest incidence rate (49%) followed by *Penicillium* (34%), each of *Cladosporium* and *Alternaria* (15%), *Acremonium* (12%), *Rhizopus* (10%), *Rhizomucor* (8%), *Absidia* (3%) and *Chrysosporium* (2%). Also, they found that five species of *Aspergillus* could be isolated from meat products samples. *A. niger* had the highest incidence rate (22%) followed by *A.flavus* (16%), *A. fumigatus* (12%), *A. parasiticus* (2%) and at least *A.ochraceus* (1%), this variation in fungal species may be due to using different types of media and storage temperature. according to [35] and [36], because of their shared ability to tolerate in low water activity (0.78-0.83), survive at low to moderate temperatures and thrive on protein-rich substrates.

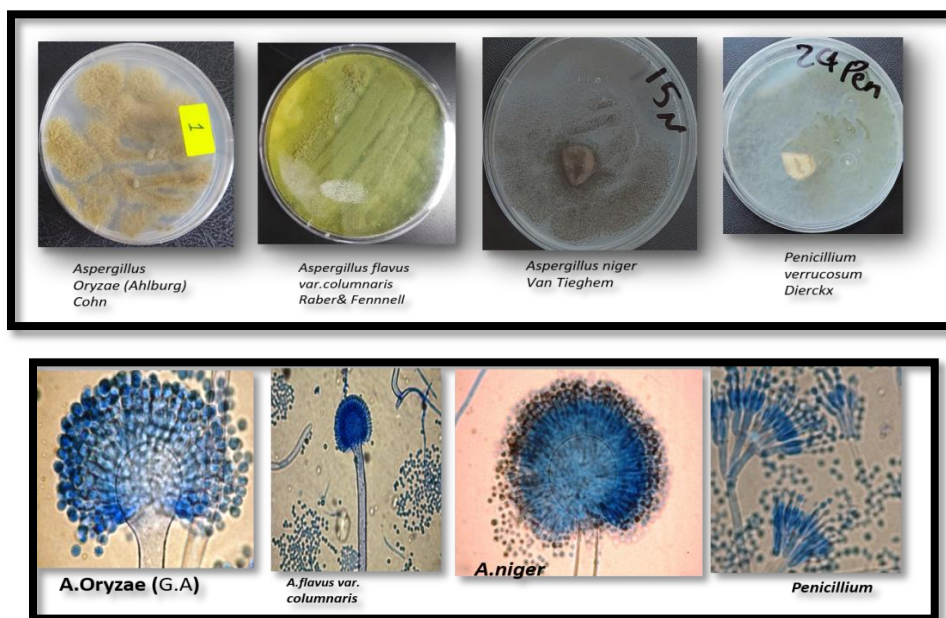
**Table2: Incidence of fungi isolated from the examined samples**

Sample/ Mold	<i>Aflavus.</i>	<i>A.niger</i>	<i>A.terrus</i>	<i>A.paraciticus</i>	<i>A.oryzae</i>	<i>A.Ochrachaeous</i>	<i>Rhizobus sp</i>	<i>Alternaria sp</i>	<i>Mucar sp</i>	<i>Trichoderma sp</i>	<i>Fusarium sp</i>	<i>Penicillium sp</i>	Total
Cornflex	4	43	0	0	0	0	0	0	1	0	1	4	53
Chicken stock	55	0	0	0	0	0	1	0	0	0	0	4	60
Meat Spices	66	62	2	0	6	1	3	0	0	0	4	9	153
Luncheon	38	248	0	2	12	0	3	0	1	0	0	9	313
Basterma	12	98	0	0	1	0	0	0	0	4	0	5	120
Burger	14	86	1	0	0	0	1	2	0	0	0	2	106
Indomie	4	10	0	0	0	0	0	0	0	0	0	0	14
Meat	0	30	0	0	0	0	0	0	0	0	0	0	30
Sausage	0	25	0	0	0	0	0	0	0	0	0	0	25
Crisps	85	66	0	0	0	0	1	0	0	0	0	0	152
Karate	5	66	0	0	0	0	1	0	0	0	0	0	72
Biscuit	42	31	0	0	0	0	0	0	0	0	0	0	73
Corn	70	0	0	0	0	0	0	0	0	0	0	0	70
Soebean	0	25	0	0	0	0	0	0	0	0	0	0	25
Total	395	790	3	2	19	1	10	2	2	4	5	33	

**Identification of fungal isolates**

AS shown in photo (1), the most prevalent isolates were determined based on preliminary morphological and cultural properties at Assiut University Mycological Centre (AUMC), Egypt with deposition numbers AUMC16127, AUMC16128, AUMC16129 and AUMC16131 for *Aspergillusoryzae*, *Aspergillusflavus* var *columnaris*, *Aspergillusniger* van tieghem and *Penicillium verrucosum*, respectively. Most of the tested meat product samples were infected with different types of fungi which considered as a major cause in the spoilage of meat products, leading to high economic losses and constitute a public health

hazard by the production of a wide variety of mycotoxins. Data shows that some tested processed meat samples, i.e. basterma, beef burger, Luncheon meat and sausage were found to be highly contaminated with fungi including *A.oryzae*, *A.flavus* var *columnaris*, *A.niger* van tieghem and *Penicillium verrucosum*. *Aspergillus* spp was the most spoilage fungi isolated from most type of foods and this result are similar to the experiments that done by Easa[40] which isolated different *Aspergillus* species from traditional fast foods. This result showed that the most important fungal species belonging to *Aspergillus flavus* and *Aspergillus niger*, which are mycotoxin-producers



**Photo (1): Fungal genera isolated from the samples**

**CONCLUSION**

Most of the examined samples including meat products as (beef, basterma, corn flex, meat spices, luncheon, burgers, indomie, meat, sausage, crisps, karate (snacks), biscuits, maize and soebean) were

contaminated with different types of fungi that regarded as a major source in food spoilage and can effect on the public health by extraction of their mycotoxins. The production of a wide variety of mycotoxins caused by the majority of fungal genera is regarded to pose serious threats to public health



and large financial losses when it comes to the deterioration of meat products.

## References

1. Shaltout F, Salem R, Fatema A, et al. Mycological evaluation of some ready to eat meat products with special reference to molecular characterization. 2016 01/01:9-14.
2. Gilbert RJ, de Louvois J, Donovan T, et al. Guidelines for the microbiological quality of some ready-to-eat foods sampled at the point of sale. PHLIS Advisory Committee for Food and Dairy Products. *Commun Dis Public Health*. 2000 Sep;3(3):163-7.
3. Sohaib M, Anjum FM, Arshad MS, et al. Postharvest intervention technologies for safety enhancement of meat and meat based products; a critical review. *J Food Sci Technol*. 2016 Jan;53(1):19-30.
4. Alcaide-Molina M, Ruiz-Jiménez J, Mata-Granados JM, et al. High through-put aflatoxin determination in plant material by automated solid-phase extraction on-line coupled to laser-induced fluorescence screening and determination by liquid chromatography-triple quadrupole mass spectrometry. *J Chromatogr A*. 2009 Feb 13;1216(7):1115-25.
5. Morshdy A, Hussien M, El-Abbasy MT, et al., editors. Aflatoxins residues in some meat products. 2nd Conference of Food Safety; 2015.
6. Navale V, Vamkudoth KR, Ajmera S, et al. Aspergillus derived mycotoxins in food and the environment: Prevalence, detection, and toxicity. *Toxicol Rep*. 2021;8:1008-1030.
7. Kumar P, Mahato DK, Kamle M, et al. Aflatoxins: A Global Concern for Food Safety, Human Health and Their Management. *Front Microbiol*. 2016;7:2170.
8. Beal T, Gardner CD, Herrero M, et al. Friend or Foe? The Role of Animal-Source Foods in Healthy and Environmentally Sustainable Diets. *The Journal of Nutrition*. 2023 2023/02/01;153(2):409-425.
9. Pereira PM, Vicente AF. Meat nutritional composition and nutritive role in the human diet. *Meat Sci*. 2013 Mar;93(3):586-92.
10. Omotayo OP, Omotayo AO, Mwanza M, et al. Prevalence of Mycotoxins and Their Consequences on Human Health. *Toxicol Res*. 2019 Jan;35(1):1-7.
11. Pandey AK, Samota MK, Kumar A, et al. Fungal mycotoxins in food commodities: present status and future concerns [Review]. *Frontiers in Sustainable Food Systems*. 2023 2023-May-05;7.
12. Soares Mateus AR, Barros S, Pena A, et al. Mycotoxins in Pistachios (*Pistacia vera* L.): Methods for Determination, Occurrence, Decontamination. *Toxins (Basel)*. 2021 Sep 25;13(10).
13. Khodaei D, Javanmardi F, Khaneghah AM. The global overview of the occurrence of mycotoxins in cereals: a three-year survey. *Current Opinion in Food Science*. 2021 2021/06/01;39:36-42.
14. Awuchi CG, Ondari EN, Ogbonna CU, et al. Mycotoxins Affecting Animals, Foods, Humans, and Plants: Types, Occurrence, Toxicities, Action Mechanisms, Prevention, and Detoxification Strategies-A Revisit. *Foods*. 2021 Jun 3;10(6).
15. Tanaka K, Sago Y, Zheng Y, et al. Mycotoxins in rice. *Int J Food Microbiol*. 2007 Oct 20;119(1-2):59-66.
16. Cheesbrough, T. M., and Kolattukudy, P. E.. Alkane biosynthesis by decarbonylation of aldehydes catalyzed by a particulate preparation from *Pisum sativum*. *Proceedings of the National Academy of Sciences*, 1984 .81(21), 6613-6617.
17. García-Linares, M. C., GONZALEZ-FANDOS, E., García-Fernández, M. C., and Garcia-Arias, M. T.. Microbiological and nutritional quality of sous vide or traditionally processed fish: Influence of fat content. *Journal of Food Quality*,2004.27(5), 371-387.
18. Thi Minh Le T, Thi Hong Hoang A, Thi Bich Le T, et al. Isolation of endophytic fungi and screening of Huperzine A-producing fungus from *Huperzia serrata* in Vietnam. *Sci Rep*. 2019 Nov 6;9(1):16152.
19. El Sayed T, El Desouky T, Abd EAAM. Investigation of fungus associated within co-occurrence of aflatoxins and ochratoxin a in cereals from Egypt. *MOJ Toxicol*. 2019;5(3):92-99.
20. Ramirez C. Manual and atlas of the Penicillia. Elsevier Biomedical Press.; 1982.
21. Pitt J, Hocking A. Fungi and food spoilage. Blackie Academic & Professional. New South Wales, Australia. 1997.
22. Ghiasian SA, Kord-Bacheh P, Rezayat SM, et al. Mycoflora of Iranian maize harvested

- in the main production areas in 2000. *Mycopathologia*. 2004 Jul;158(1):113-21.
23. Pitt JI, Hocking AD. *The Ecology of Fungal Food Spoilage*. Fungi and Food Spoilage. Boston, MA: Springer US; 2009. p. 3-9.
  24. El-Kady I, Zohri A, editors. *Mycoflora and mycotoxins of beef burger in Egypt*. 1st Symposium on Food Safety; 2000.
  25. Zohri A, Moharram A, Refaie R. Mycobiota contaminating beef burger and sausage with reference to their toxins and enzymes. *J Basic Appl Mycol (Egypt)*. 2014;5:61.
  26. Omorodion, N. J. P. N., and Odu, N. N.. Microbiological quality of meats sold in Port Harcourt Metropolis, Nigeria. *Nat Sci*,2014. 12(2), 58-62.
  27. Ismail S, Shehata A, El-Diasty E. Microbiological quality of some meat products in local markets with special reference to mycotoxins. *Global Veterinaria*. 2013;10(5):577-584.
  28. Nabil GN, Khafaga NI, El-Hariri M, et al. Detection of fungi and total aflatoxins in food additives and some meat products by serological and molecular biological methods. *Journal of the Egyptian Veterinary Medical Association*. 2017;177(2):153-173.
  29. Elgazzar M, Abdo A, El-Zeny M. Mycological assessment of cooked beef products. *Mansoura Veterinary Medical Journal*. 2019;20(2):12-19.
  30. Chehri K, Azami E, Mosaber A. *Aspergillus* and aflatoxin B1 contamination of stored corn grains in western Iran. *Global Veterinaria*. 2015;14(1):39-42.
  31. Bhat RV, Shetty PH, Amruth RP, et al. A foodborne disease outbreak due to the consumption of moldy sorghum and maize containing fumonisin mycotoxins. *J Toxicol Clin Toxicol*. 1997;35(3):249-55.
  32. Gao J, Liu Z, Yu J. Identification of *Aspergillus* section *Flavi* in maize in northeastern China. *Mycopathologia*. 2007 Aug;164(2):91-5.
  33. Fatema A. Mycological evaluation of some ready to eat meat products with special reference to molecular characterization\* Shaltout, FA;\*\* Salem, R. M;\*\* El-diasty, Eman and. *Veterinary Medical Journal–Giza*. 2016;62(3):9-14.
  34. Kocić-Tanackov SD, Dimić GR, Karalić D. Contamination of spices with moulds potential producers of sterigmatocystine. *Acta Periodica Technologica*. 2007 (38):29-35.
  35. Abdeltawab A, M E-D, Khater D, et al. Mycological Identification of Some Fungi Isolated from Meat Products and Spices with Molecular Identification of Some *Penicillium* Isolates. *Advances in Animal and Veterinary Sciences*. 2020 01/18;8.
  36. Shaltout F, Salem R. Moulds, aflatoxin B1 and ochratoxin A in frozen livers and meat products. *Veterinary Medical Journal Giza*. 2000;48(3):341-346.
  37. Brr A, Moustafa N, Edris A. Incidence of moulds and aflatoxins in some meat products. *Benha Vet Med J*. 2004;15(2):65-75.
  38. Filtenborg O, Frisvad JC, Samson R. Specific association of fungi to foods and influence of physical environmental factors. *Introduction to food-and airborne fungi*. 2004 (Ed. 7):306-320.
  39. Frisvad JC, Skouboe P, Samson RA. Taxonomic comparison of three different groups of aflatoxin producers and a new efficient producer of aflatoxin B1, sterigmatocystin and 3-O-methylsterigmatocystin, *Aspergillus rambellii* sp. nov. *Syst Appl Microbiol*. 2005 Jul;28(5):442-53.
  40. Easa SMH. The microbial quality of fast food and traditional fast food. *Nature and Science*. 2010;8(10):117-133.